

Clinical Commentary Review

Practical Management of Antibiotic Hypersensitivity in 2017

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Antibiotics are the most common class of medications that individuals report allergy or intolerance to. Adverse reactions are reported at a predictable rate with all antibiotic use that vary by antibiotic. Antibiotic allergy incidence rates are sex dependent, higher in females than in males. Most of these events are not reproducible or immunologically mediated. Antibiotic allergy prevalence increases with increasing age and is more common in hospitalized populations and in populations that use more antibiotics. Determining potential mechanisms for the observed symptoms of the adverse reactions is the starting point for effective management of antibiotic hypersensitivity. Skin testing and direct challenges are the primary tools used to determine acute tolerance in 2017. Commercially available *in vitro* testing is not currently clinically useful in determining antibiotic hypersensitivity, with rare exceptions. Desensitization can be used when acute-onset immunologically mediated hypersensitivity is confirmed to safely administer a needed antibiotic. Desensitization is not possible when clinically significant T-cell-mediated delayed-type hypersensitivity is present. Effective management of antibiotic allergy is an important part of a comprehensive antibiotic stewardship program. © 2017 American Academy of Allergy, Asthma & Immunology (J Allergy Clin Immunol Pract 2017;■:■-■)

Key words: Allergy; Antibiotic; Challenge; Desensitization; Hypersensitivity; intolerance; Management; Skin test

Why another review on antibiotic hypersensitivity? Why would you want to reexpose an individual to an antibiotic they think already gave them a problem? We all know that antibiotics

are significantly overused and the pattern of overuse varies widely across countries and across continents, wherever antibiotics are available.^{1,2} We also recognize that there are no side effect-free drugs and that all antibiotic use is associated with a certain predictable rate of adverse reactions. Multiple drug-resistant infections are becoming more common.³ Developing effective antibiotic stewardship programs is taking on new importance and urgency.⁴ New information on the risks of avoidance of antibiotics, particularly narrow-spectrum penicillins, when they are the drugs of choice has made it important for specialists in Allergy and Clinical Immunology and Infectious Disease along with hospitalists and other primary care providers to be able to safely and efficiently evaluate and/or refer individuals with reported antibiotic intolerances and know when to do diagnostic testing, drug challenge, or desensitize.⁵

This review is meant to provide a practical approach to managing the clinical care of individuals who have reported adverse reactions to antibiotics. In this article, we use the term “intolerance” to encompass any reaction that is typically listed as an *allergy* in the medical record, recognizing that most of these are not true immunologic reactions and many may be perceived symptoms unrelated to the drug. The literature on evaluating antibiotic-associated adverse drug reactions has evolved dramatically over the past 80 years, since sulfonamide antibiotics became clinically available. The cumulative literature is very confusing, often apparently contradictory, and almost impossible to reconcile in its entirety.⁶⁻⁸ Nonimmunologically mediated reactions are commonly referred to as “*allergy*” in the electronic health record (EHR), which adds to the confusion in both patients and many physicians.⁹ Given this, it is not surprising that current antibiotic hypersensitivity management and testing practices vary widely.¹⁰

Most reported antibiotic intolerance or *allergy* is from direct pharmacologic effects of the antibiotic or antibiotic-independent sequela of the underlying bacterial or viral infection being treated, or often mistreated, with the implicated antibiotic.¹¹ These nonimmunologically mediated sequelae can include headaches, fevers, benign rashes, hives with or without angioedema, gastrointestinal symptoms including upset stomach, nausea, and vomiting, serious drug-resistant infections including methicillin-resistant *Staphylococcus aureus*, vancomycin-resistant enterococcus, and *Clostridium difficile*, and other somatic symptoms.¹² Antibiotic-associated immunologically mediated hypersensitivity only accounts for a small subgroup of reported intolerances or *allergy*.¹¹ Immunologically mediated symptoms can vary from relatively common, but benign, hives or delayed-onset exanthems, to serious and potentially life-threatening reactions such as IgE-mediated anaphylaxis, serious cutaneous adverse drug reactions (SCARs), hepatitis, serum sickness,

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Abbreviations used*EHR- Electronic health record**DRESS- Drug eruption with eosinophilia and systemic symptoms**SCARs- Serious cutaneous adverse reactions**SJS- Stevens-Johnson syndrome**SSLR- Serum sickness-like reaction**TEN- Toxic epidermal necrolysis*

hemolytic anemia, cytopenias, pneumonitis, or nephritis.¹¹ We will provide a general framework on how to address the management of individuals presenting with antibiotic *allergy*, safely identify those with clinically significant immunologically mediated hypersensitivity, and enable most of them to use the needed antibiotic or the least morbid alternative.

EPIDEMIOLOGY OF ANTIBIOTIC HYPERSENSITIVITY

The reference standard for a clinically significant antibiotic-associated adverse event is currently what is reported in the drug *allergy* field of the EHR.¹³ In most EHRs there is only a drug *allergy* field and no other place to note drugs that should not be used, used with caution, or were associated with an adverse reaction. If there is no notation made in the EHR, then any adverse reaction that may have occurred cannot affect future antibiotic choice, and an inaccurate notation, or unconfirmed *allergy*, in the EHR will greatly reduce future exposure to that antibiotic. When one reviews the EHR, the time after the antibiotic exposure can make a significant difference in the adverse reaction incidence rate observed. Not all reports are made in a timely fashion. Checking 1 month after an exposure results in significantly lower reported rates of antibiotic *allergies* than checking 1 year after.¹⁴ There are limited comprehensive population-based data on the incidence and prevalence of reported antibiotic intolerances (*allergies*). The prevalence of drug intolerance reports varies around the world, apparently dependent on the most commonly used antibiotics. In a study of 2,375,424 Kaiser Permanente Southern California healthplan members, who had at least 11 months of coverage in 2009, at least 1 visit during 2009, and essentially a representative sample of about 1% of the US population that uses health care, the prevalences of *allergy* to specific antibiotics were as follow: penicillins 186,630 (7.9%), sulfonamides 101,952 (4.3%), macrolides 28,275 (1.2%), cephalosporins 25,956 (1.1%), tetracyclines 16,555 (0.70%), quinolones 18,815 (0.46%), nitrofurantoin 5,809 (0.24%), clindamycin 4,745 (0.20%), metronidazole 3,623 (0.15%), with only rare reports to other antibiotics.¹⁵ See [Table I](#). There was a marked female predominance for all reported antibiotic intolerances, with the highest ratios noted for nitrofurantoin (0.43% females, 0.02% males, ratio, 21.5) and metronidazole (0.25% females, 0.04% males, ratio, 6.25). Overall, there were 216,192 (16.7%) females who reported at least 1 antibiotic intolerance, compared with only 94,576 (8.8%) of the males. Similar data were obtained from 1,766,328 hospitalized patients in Boston between 1990 and 2013, with the expected higher rates of drug intolerances noted in relatively older hospitalized patients compared with the entire population.¹⁶ In contrast, in a study of an academic general practice in the

Netherlands in 2014, less than 2% of the 8288 patients reported penicillin intolerance.¹⁷

New antibiotic intolerance reporting incidence is higher in females than in males for all antibiotic families.¹⁸ It is also higher for all antibiotic families, except penicillins, in individuals with any reported drug intolerance compared with individuals with no reported drug intolerances. About 1 in 200 penicillin antibiotic courses results in a new penicillin intolerance report, independent of drug intolerance history.¹⁵ Individuals receiving penicillins after negative penicillin allergy testing note reactions about 1% to 3% of the time, but only rarely convert to allergy test positive.¹⁹ Individuals with multiple drug intolerance syndrome rarely have any underlying immunologically mediated mechanism responsible for their reactions because most listed reactions are predictable side effects.²⁰

CLINICAL PRESENTATIONS

Clinically, hypersensitivity reactions to antibiotics are classified as immediate and nonimmediate according to the time interval between the first dose of the last administration and their onset.^{21,22} *Immediate reactions* are defined as those occurring within 1 hour and up to 6 hours after the first dose of the last administration.^{21,23} Immediate reactions usually manifest as isolated symptoms, such as urticaria, angioedema, conjunctivitis, respiratory symptoms (rhinitis, bronchospasm, cough, dyspnea), gastrointestinal symptoms (nausea, vomiting, diarrhea, abdominal pain), or rarely as anaphylaxis.²² *Nonimmediate reactions* are defined as those occurring at any time greater than 6 hours after the initial drug administration,²² often starting 2 to 5 days later. The most common nonimmediate reactions are maculopapular or morbilliform exanthems and delayed-appearing urticaria/angioedema. Rarely, SCARs occur, which include acute generalized exanthematous pustulosis, drug-induced hypersensitivity syndrome or drug rash (or reaction) with eosinophilia and systemic symptoms (drug eruption with eosinophilia and systemic symptoms [DRESS]), and severe bullous exanthems such as Stevens-Johnson syndrome (SJS) and toxic epidermal necrolysis (TEN).²⁴ In addition, nonimmediate reactions can be manifest by interstitial nephritis, pneumonitis, hemolytic anemia, cytopenias, hepatitis, and/or vasculitis with or without signs of serum sickness occurring up to several weeks after the first dose of the last administration.

If there is a recent history of antibiotic-associated anaphylaxis, specifically within 3 months of testing, the probability of clinically significant IgE-mediated allergy will be high, 20% to 50%.²⁵ If there is a distant history of benign index symptoms, specifically more than 10 years ago, the risk of clinically significant IgE-mediated allergy will be very low, 1% to 2%.²⁶ For individuals with a distant history of anaphylaxis, the risk is lower than for more recent reactions and many reported episodes of anaphylaxis do not meet the clinical criteria for true anaphylaxis, making the repeat risk even lower. Testing strategies need to consider the pretest probability of a positive test result. There are also frequent false-positive test results for most skin testing protocols and *in vitro* testing methods.²⁷ If these are relied on solely to determine acute tolerance and the reference standard of a drug challenge is not used to confirm, then many more individuals will suffer morbidity from the inability to use the preferred antibiotic they could safely tolerate.

TABLE I. Intolerance prevalence and immunogenicity of common antibiotics

Antibiotic or antibiotic family	Intolerance prevalence	IgE-mediated allergy	T-cell-mediated delayed hypersensitivity	Intrafamily immunologic cross-reactivity
Penicillins	7.9%	Possible	Possible	Common
Sulfonamides	4.3%	Unlikely	Possible	Unlikely
Macrolides	1.2%	Unlikely	Unlikely	Unknown
Cephalosporins	1.1%	Possible	Possible	Unlikely
Tetracyclines	0.70%	Unlikely	Unlikely	Unknown
Quinolones	0.46%	Possible	Unknown	Common
Nitrofurantoin	0.24%	Unlikely	Unlikely	NA
Clindamycin	0.20%	Unlikely	Possible	NA
Metronidazole	0.15%	Unlikely	Possible	NA

NA, Not applicable/available.

MECHANISMS OF IMMUNOLOGICALLY MEDIATED ANTIBIOTIC HYPERSENSITIVITY

The immunologic mechanism involved in most immediate reactions is classically antigen-specific IgE-dependent mast cell activation. It is also possible for antigen-specific IgG to activate complement that results in acute rashes and non-IgE-mediated mast cell activation. Nonimmediate reactions are even more heterogeneous. Most confirmed immunologically mediated maculopapular exanthems are T-cell-mediated. Cytotoxic and IL-5-producing, drug-specific CD4 T cells migrate into the skin and kill keratinocytes presenting the drug on MHC class II molecules through a perforin-dependent mechanism.^{24,28} SCARs are frequently mediated through antigen-specific T cells.²⁹ Antigen-specific IgG and complement can be responsible for reactions as varied as delayed serum-sickness reactions, hemolytic anemias and cytopenias, and anaphylactic reactions.³⁰ It is possible that a prolonged antibiotic course results in the new production of antibiotic-specific IgE and results in delayed-onset urticaria.³¹

ELEMENTS FOR PROPER DOCUMENTATION OF DRUG INTOLERANCES/ALLERGY

Table II outlines the 3 essential elements of a drug intolerance entry in the EHR: the date of the reaction, the time between the first dose of the last course and the onset of the symptoms, and the specific symptoms.³² There needs to be enough information listed in the drug intolerance entry to enable future treating physicians to determine whether any of the following is possible or probable: IgE-mediated allergy, IgG and complement-mediated hypersensitivity, T-cell-mediated delayed-type hypersensitivity, or any SCAR (including DRESS, TEN, SJS, or acute generalized exanthematous pustulosis) or serum sickness-like reaction (SSLR).

DIAGNOSIS AND TESTING CONCERNS RELATED TO SPECIFIC ANTIBIOTICS OR ANTIBIOTIC FAMILIES

Penicillins

Needless avoidance of penicillin because of an unconfirmed penicillin allergy has been shown to be a significant public health risk.^{33,34} Any effective testing strategy needs to rapidly, inexpensively, and safely test individuals and minimize false-positive test results.³⁵ There is a significant level of IgE-mediated immunologic cross-reactivity among penicillins based on the

TABLE II. The 3 essential elements of a drug intolerance entry in the EHR

1. Date of the index adverse reaction
2. Time to onset of the index adverse reaction after the first dose of the last course
3. Specific clinical characteristics of the index adverse reaction (note 1 or more of the following in as much detail as possible)
a. Anaphylaxis or shortness of breath*
b. Hives*
c. Other benign rashes*
d. Gastrointestinal symptoms*
e. Other benign symptoms*
i. Headaches
ii. Cough
iii. Other
iv. Unknown
f. SCARs or other serious systemic symptoms‡
i. Biopsy proven T-cell-mediated severe cutaneous eruption
ii. DRESS
iii. SJS
iv. TEN
v. Hepatitis/pneumonitis/nephritis
vi. Hemolytic anemia/cytopenias
vii. Other severe nonimmunologically mediated symptoms
A. Angioedema
B. Tendon rupture
C. Other

*OK to consider testing, drug challenge, or desensitization if indicated.

‡Do not consider testing or drug challenge outside of a research protocol. Desensitization is not possible.

shared core structures.³⁶ Aminopenicillins typically have higher rates of IgE-mediated adverse reactions compared with other penicillins.³⁷ There are individuals specifically reactive only to amoxicillin, a phenomenon apparently more common in Europe than in North America.^{38,39} The reference standard test to document acute penicillin class antibiotic tolerance is an oral challenge, usually with amoxicillin, the most commonly used penicillin.⁴⁰ Low-risk individuals, including those with benign delayed-onset nonurticarial rashes, gastrointestinal upset, or headaches, can be safely evaluated with direct oral challenge without any preliminary skin testing.⁴¹ Skin testing is done to minimize the number of serious acute oral challenge reactions. There is no consensus approach to what testing reagents are necessary or sufficient when penicillin skin testing, but simpler protocols appear to be safe and effective in most clinical settings.

In North America, where penicillin minor determinants are not commercially available, skin testing with benzylpenicilloyl-polylysine (PRE-PEN) and penicillin G, and when negative followed by a single dose of amoxicillin 250 mg, has been shown to be a safe and effective method for determining tolerance to penicillin.⁴² Penicillins can also induce T-cell-mediated delayed hypersensitivity reactions.⁴³ Delayed reading of intradermal testing or patch testing has been used to try to predict clinically significant T-cell-mediated reactions, but results have not been consistent and the predictive value is uncertain; therefore, an oral challenge remains the reference standard test.⁴⁴ Multiday challenges do not appear to be any safer or clinically more useful than single-day challenges.⁴⁵ If there is a clinically significant T-cell-mediated, non-SCAR, delayed-type hypersensitivity to amoxicillin, it will typically be manifest within 5 days of an oral 250 mg amoxicillin challenge.⁴⁶ *In vitro* antipenicillin IgE tests are not useful outside of the setting of evaluating individuals who recently had penicillin-associated anaphylaxis.^{47,48} When both skin tests and *in vitro* tests are highly positive, the risk of repeat anaphylaxis with an oral challenge is high.⁴⁹ Those rare individuals with penicillin-associated SCARs should not be tested or rechallenged outside of a research setting.⁵⁰

Sulfonamide antibiotics

Sulfonamide antibiotics (eg, sulfamethoxazole, sulfadoxin, and sulfapyridine) are sulfonyl arylamines, characterized by a sulfonamide (SO₂-NH₂) moiety directly attached to a benzene ring, which carries an unsubstituted amine (-NH₂) at the N₄ position.^{51,52} There is no cross-reactivity between antibiotic and nonantibiotic sulfonamides. Sulfonamides are most commonly associated with nonimmediate manifestations, such as maculopapular rashes and fixed drug eruptions. SCARs have been reported.^{52,53} The risk of SJS/TEN is higher for sulfonamide antibiotics than for most other antibiotics. Most immunologically mediated sulfonamide-associated adverse reactions appear to be T-cell-mediated.⁵⁴ Sulfonamide antibiotics have the highest benign rash rate of all antibiotics.¹⁵ The rash rate is even higher in individuals with active untreated or acutely treated HIV infection with low CD4 T-cell counts.⁵⁵ Prolonged treatment of the HIV with highly effective antiretroviral therapy then often results in tolerance.⁵⁶ There is no useful skin or *in vitro* test available to gauge the acute risk of reexposure.⁵⁷ Patch testing and the delayed reading of intradermal tests have been used to help gauge the delayed-onset risk of reexposure, but clinically drug challenge is still the reference standard test.⁵⁸ Dapsone does not appear to significantly cross-react with sulfamethoxazole.⁵⁹ Single-dose challenges appear to be just as safe as multiple dose challenge/desensitizations for proving tolerance to sulfonamides in HIV-infected patients.^{60,61} Whether this is true for non-HIV patients is unknown but multistep single-day challenges have been shown to be as safe and effective as multistep multiday challenges.⁵⁸

Macrolides

Macrolides are classified according to the number of carbon atoms in their lactone ring: 14 membered (erythromycin, troleandomycin, roxithromycin, dirithromycin, and clarithromycin), 15 membered (azithromycin), and 16 membered (spiramycin, rokitamycin, josamycin, and midecamycin).⁶² Azithromycin is the most widely overused antibiotic in the United States.⁶³ Macrolide antibiotics reversibly bind bacterial 50S

ribosomal subunits. They are too small to be directly immunogenic. It is unknown whether they are able to haptenate serum proteins. They are promotility agents and can be directly irritating to the gastrointestinal tract. Data on the utility of skin testing with macrolides is conflicting and although several studies have evaluated clarithromycin skin tests using a lyophilized form, this is not available in the United States. Because of several potential reasons including their direct irritant effect on the skin, their small size, and their lack of covalently binding to serum proteins, on the basis of the available evidence, the authors believe that macrolide skin testing is generally not useful due to the potential for false-positive skin test results at concentrations below the published nonirritating concentrations.^{64,65} There does not appear to be any clinically significant class intolerance noted with macrolides.⁶⁶ If one macrolide is not tolerated, select another if indicated. If a specific macrolide is needed again, a drug challenge is safe and effective.^{66,67}

Cephalosporins

About 3% of individuals exposed to a third- or higher-generation cephalosporin develop *Clostridium difficile* infection within 90 days.⁶⁸ First- and second-generation cephalosporins have *Clostridium difficile* infection incidence rates an order of magnitude lower.⁶⁸ Cephalosporins in general do not have clinically significant levels of immunologic cross-reactivity with penicillins, except for rare individuals who mount specific responses to shared side chains.⁶⁹ Penicillin allergy testing is not useful in evaluating the risk of cephalosporin-associated reactions.⁷⁰ Individuals with a history of intolerance to one cephalosporin typically tolerate other cephalosporins.⁷¹ Skin testing and *in vitro* testing are not well enough validated, or even needed, for routine use in the clinical management of cephalosporin allergy.⁷² Typically, just selecting an alternative cephalosporin, which ideally does not share a side chain with the implicated cephalosporin, is a safe and commonly used approach.⁶⁸ In patients with anaphylaxis to cephalosporins, negative skin tests to cephalosporins with disparate R1 groups predicted tolerance.⁷² Cephalosporins are only extremely rarely uniquely associated with SCARs.⁶⁸

Tetracyclines

Tetracyclines can induce photosensitive dermatitis and esophageal irritation.⁷³ They have not been convincingly shown to induce clinically significant IgE-mediated allergy or T-cell-mediated hypersensitivity. Drug challenge is generally successful.

Quinolones

Quinolones can be classified according to their generation: first (cinoxacin and nalidixic acid), second (ofloxacin, norfloxacin, ciprofloxacin, and enoxacin), third (levofloxacin), and fourth (gemifloxacin and moxifloxacin). Clear patterns of immunologically mediated cross-reactivity among quinolones have not been well established.⁷⁴ Skin testing with quinolones is not clinically useful because of direct mast cell activation through the Mrgprb2 receptor.^{75,76} Levofloxacin-mediated histamine release is closely linked to activation of pertussis toxin-sensitive G proteins.⁷⁷ IgE-mediated hypersensitivity has been suggested using *in vitro* testing. Apparent T-cell-mediated SCARs have been rarely identified. Clinically the reference standard test for acute tolerance remains drug challenge. Nonimmunologically mediated adverse reactions predominate. Both hypoglycemia and

TABLE III. Suggested drug challenge protocols based on risk factors

Risk factors	Direct full-dose challenge (oral if possible, 1 h of observation after dose if history of acute reaction and 5 d of follow-up if history of delayed-onset reaction)	Graded challenge (1/10th and then a full dose with 1/2 h of observation after first dose and 1/2-1 h of observation after the final dose)	Graded challenge (1/100th, 1/10th, and then a full dose with 1/2 h of observation after the first 2 doses and 1/2-1 h of observation after the final dose)
Low-probability medication	X		
Hives or other benign index reaction			
Low-probability medication		X	
Possible anaphylaxis			
High-probability medication		X	
Hives or other benign index reaction			
High-probability medication			X
Comorbid disease (eg, severe chronic obstructive pulmonary disease)			

TABLE IV. Penicillin oral desensitization protocol⁹⁹

Step	Penicillin V (units/mL)	Dose (mL)	Dose (units)	Cumulative dose
1	1000	0.1	100	100
2	1000	0.2	200	300
3	1000	0.4	400	700
4	1000	0.8	800	1,500
5	1000	1.6	1600	3,100
6	1000	3.2	3200	6,300
7	1000	6.4	6400	12,700
8	10,000	1.2	12,000	24,700
9	10,000	2.4	24,000	48,700
10	10,000	4.8	48,000	96,700
11	80,000	1.0	80,000	176,000
12	80,000	2.0	160,000	336,700
13	80,000	4.0	320,000	656,700
14	80,000	8.0	640,000	1,296,700

Administer penicillin V orally every 15 min per step. Each dose can be diluted in 30 mL water for patient to ingest. Total time: 3 h 45 min.

hyperglycemia have been noted among elderly hospitalized patients who received gatifloxacin.⁷⁸ Quinolones have been associated with tendon rupture.⁷⁹

Nitrofurantoin

Nitrofurantoin has not been convincingly associated with IgE- or benign T-cell-mediated reactions. Serious nitrofurantoin-induced pulmonary toxicity is well described and rechallenge is contraindicated. Reproducible nitrofurantoin-induced hepatic injury is also well described and rechallenge is contraindicated. Recurrence has been documented as long as 17 years after the index reaction.⁸⁰ Benign rashes can be safely rechallenged.

Clindamycin

Clindamycin is associated with much higher rates of *Clostridium difficile* infections than are penicillins.⁸¹ Clindamycin

has been associated with T-cell-mediated delayed-onset hypersensitivity that may be predicted by patch testing.⁸²

Metronidazole

Metronidazole has not been convincingly associated with IgE- or benign T-cell-mediated reactions. Drug challenge is generally successful.⁸³

Other betalactams

Other betalactams, including monobactams, carbapenems, and clavulanate, do not tend to have clinically significant levels of immunologic cross-reactivity with penicillins or cephalosporins, with the exception of rare individuals who mount specific responses to shared side chains.⁸⁴⁻⁸⁸ Individuals with IgE-mediated allergy to betalactamase inhibitors such as clavulanate have been identified.⁸⁹

Aminoglycosides

Aminoglycosides are classified into 2 groups: the streptidine group (streptomycin) and the desoxystreptamine group (kanamycin, amikacin, gentamicin, tobramycin, and neomycin).⁹⁰ Aminoglycosides can rarely cause both immediate and non-immediate hypersensitivity reactions. Skin testing and patch testing have been rarely used. Cross-reactivity among aminoglycosides is common, approaching 50% or more among those belonging to the desoxystreptamine group.⁹¹ Aminoglycosides are directly nephrotoxic and ototoxic.

Vancomycin

Vancomycin can directly activate mast cells and is associated with a "red man syndrome." Premedication with antihistamines, slowing the infusion rate, and avoidance of concomitant mast cell secretagogues (eg, opiates) are helpful in patients with red man syndrome. Vancomycin has been associated with rare cases of T-cell-mediated delayed-type hypersensitivity, linear IgA bullous dermatosis, DRESS, acute interstitial nephritis, and SJS/TEN.^{92,93} Patch testing and skin testing have been used to help gauge the risk of reexposure.⁹⁴

TABLE V. Example intravenous drug desensitization protocol

Preparation of solutions					
	Volume of diluent (eg, 0.9% sodium chloride)		Total amount to be injected in each solution		Final concentration
Solution 1	250 mL		10 mg		0.04 mg/mL
Solution 2	250 mL		100 mg		0.4 mg/mL
Solution 3	250 mL		1000 mg		4 mg/mL
Drug desensitization protocol					
Step	Solution	Rate (mL/h)	Time (min)	Administered dose (mg)	Cumulative dose (mg)
1	1	2	15	0.02	0.02
2	1	5	15	0.05	0.07
3	1	10	15	0.1	0.17
4	1	20	15	0.2	0.37
5	2	5	15	0.5	0.87
6	2	10	15	1	1.9
7	2	20	15	2	3.9
8	2	40	15	4	7.9
9	3	10	15	10	18
10	3	20	15	20	38
11	3	40	15	40	78
12	3	80	15	80	158
13	3	160	15	160	318
14	3	320	15	320	638
15	3	640	9	362	1000

Total time = 219 min.

Full dose = 1000 mg.

DRUG CHALLENGES

Drug challenges are considered the reference standard for confirming acute tolerance to an antibiotic. Even in the case of penicillin allergy where skin testing has a good negative predictive value, an oral challenge is recommended to confirm acute tolerance and convince the patient that the antibiotic can be safely used. For most antibiotics, skin testing is not as well standardized and clinically useful and drug challenges are relied on to confirm tolerance. Many terms are used to describe drug challenge procedures including graded dose challenges, test doses, and provocation. The term provocation implies that the intention of this procedure is to provoke a reaction when in fact the purpose is quite the opposite. In clinical practice, the purpose of drug challenge is to confirm tolerance to an antibiotic and in carefully selected patients serious reactions are uncommon.^{95,96} Pretreatment with antihistamines is not generally recommended for antibiotic drug challenge.

A negative drug challenge typically indicates that the patient may be considered acutely tolerant to that antibiotic. However, false-negative drug challenges may occur because of several factors including inadequate dosing, inadequate duration of the drug challenge, or a missing cofactor (eg, no concurrent infection). There is also a certain rate of adverse reactions that occur with all antibiotic use, even in individuals with no previous reactions. In the case of betalactam antibiotics, false-negative results occur infrequently as evidenced by a report by Demoly et al⁹⁷ that determined a negative predictive value of 94%. Of these false-negative reactions, none was serious, most were

delayed, and all were cutaneous (mostly urticaria or exanthems) and some patients with self-reported reactions had subsequent negative drug challenges.

In clinical practice, drug challenges for antibiotic allergy are intended for patients who are deemed to be at low risk for being allergic to the given drug.⁹⁸ They are typically performed after skin testing, such as the case for penicillin, or may be performed without skin testing. Nonirritating concentrations are not well established for quinolones and some antibiotics may be irritating even at very dilute concentrations; therefore, drug challenges remain the only option for the initial evaluation of a patient. Challenges may also be used to exclude cross-reactivity. For example, most IgE-mediated cephalosporin allergy is apparently directed against R1-specific side-chain determinants. Patients who have negative skin tests to other cephalosporins with disparate R1 side chains have been shown to tolerate drug challenges with cephalosporins.⁹⁹ Many patients may have vague or subjective symptoms that are mild and not convincing for an IgE-mediated drug allergy (eg, headache or nausea) and a drug challenge can help confirm tolerance of these individuals. Drug challenges are contraindicated in antibiotic-induced lupus, vasculitis, organ-specific drug reactions causing cytopenias, hepatitis, nephritis, or pneumonitis and severe cutaneous adverse reactions including SJS and DRESS. Drug challenges are generally avoided in patients with SSLR; however, there are some data suggesting that children with SSLR to amoxicillin may tolerate amoxicillin challenges.^{41,100} Whether this approach is safe for other antibiotics has not been studied. However, it should be noted that SSLRs to cefaclor are typically

cefactor-specific and these patients tolerate other cephalosporins with no greater risk than average patients.¹⁰¹

There are multiple published protocols for antibiotic challenges, all with some fairly common approaches. Drug challenges for patients with immediate reactions should always be performed in a setting capable of treating anaphylactic reactions, although these are rarely encountered with good patient selection criteria. Determining the starting dose for an antibiotic challenge depends primarily on the pretest probability of the patient being truly allergic to that antibiotic.¹⁰² See Table III. The key is using at least a 10-fold dose increment during multidose drug challenges. The lower the likelihood the patient will have a serious acute reaction, the higher the starting dose. Typical starting doses range from 1/100th of the final dose to a full therapeutic dose. More than 3 steps are virtually never indicated. For patients with histories of immediate reactions, doses are commonly administered every 30 to 60 minutes and typically patients are observed for 1 hour following the final dose. Drug challenges using these approaches have been found to be safe and effective methods for determining tolerance. If an acute IgE-mediated reaction occurs during a drug challenge, desensitization should be performed if that antibiotic is needed in the future.

Although some allergists use multistep challenges involving 3 or more graded doses, when compared with either a single-step or a 2-step challenge, reaction rates and severity are similar and there appears to be no advantage to these more expensive and time-consuming approaches.¹⁰² Furthermore, using multiple steps, particularly with less than 10-fold dosing increments, may inadvertently desensitize the patient, which is not the goal of a drug challenge.

For delayed reactions, a single therapeutic dose with 2 to 5 days of home follow-up is generally sufficient. Patients can report the reactions and then return to the clinic for observation, treatment, and/or a skin biopsy if needed. The drug can be readministered every several days to allow time for a reaction to develop, if there is concern. For patients with a history of non-immediate benign cutaneous reactions, challenges can be performed at home, but the first dose may be given under supervision if there is uncertainty.

For patients with purely subjective symptoms or with a history of multiple drug intolerance syndrome, one should strongly consider performing single-blind placebo-controlled challenges because these patients frequently have symptoms with placebo.¹⁰² For oral challenges, inert fillers such as methylcellulose can be inserted into opaque gelatin capsules for use as placebos.

ANTIBIOTIC DESENSITIZATION

Antibiotic desensitizations have been performed since the 1940s, initially with gradually increasing doses of penicillin concomitant with antihistamine and epinephrine injections.¹⁰³ However, it was not until the early 1980s when a safer desensitization protocol was developed by Sullivan et al¹⁰⁴ and most antibiotic desensitization protocols are quite similar to this. See Table IV. Oral desensitizations should be performed whenever possible, even before the parenteral use of the same antibiotic. The basic principle of an antibiotic desensitization is to start with about double the dose used to produce the positive skin test result that confirmed the presence of clinically significant amounts of antibiotic-specific IgE. This is generally about 1/10,000th of the final therapeutic dose, with doubling doses every

15 minutes. If a reaction occurs during the desensitization, treat the reaction, stabilize the patient, then repeat the dose that was last tolerated, and complete the desensitization. Desensitizations can be done even in pregnant patients.¹⁰⁵ Castells¹⁰⁶ has developed a protocol using 3 different dilutions of a parenteral antibiotic starting with a 1:100 dilution, then a 1:10 dilution, and then the full concentration with advances in dosing made by increasing the rate of infusion¹⁰⁶ (Table V). Adverse reactions to antibiotic drug desensitizations are generally less frequent, and no more severe, than with other desensitizations. They can be safely performed in an outpatient setting that routinely performs oral food challenges. Individuals performing antibiotic desensitizations need to be well trained and comfortable in the management of acute anaphylaxis. In one of the early studies by Sullivan et al¹⁰⁴ using oral penicillin desensitizations, about 30% had delayed-onset reactions, none had immediate or anaphylactic reactions. Desensitization should be attempted only in the setting of well-documented IgE-mediated antibiotic allergy. Desensitizations are not clearly effective for T-cell-mediated reactions, such as benign delayed-onset rashes, and are contraindicated for more serious T-cell-mediated reactions.¹⁰⁷ Desensitizations are not possible for other nonimmunologically mediated reactions.¹⁰⁸

MAINTENANCE OF ACUTE TOLERANCE AFTER DESENSITIZATION

It is critical to have continual exposure to the antibiotic after desensitization until the therapeutic course is completed. If more than 5 half-lives of the antibiotic expire without further dosing, then it will be necessary to repeat the desensitization. In certain settings, reactions can be noted with delayed redosing after as little as 2 half-lives, but reactions are also common in desensitized individuals even with continual dosing. In the setting of long-acting benzathine penicillin used for syphilis therapy, patients appear to be capable of maintaining a desensitized state for as long as 3 weeks and repeat desensitization is typically not needed.¹⁰⁴

CONCLUSIONS

The effective management of antibiotic hypersensitivity starts with the accurate documentation of the index reaction. It is necessary to determine the likely mechanism(s) compatible with the clinical symptoms observed. After a thorough evaluation, the vast majority of patients with reported antibiotic allergy can be shown to be tolerant of the drug. Perform skin testing or *in vitro* testing when indicated and/or possible to screen out individuals at high risk for IgE-mediated anaphylaxis. Drug challenge, orally when possible, remains the reference standard test to determine acute tolerance. New reactions are always possible with repeat therapeutic administrations, even in drug challenge negative individuals. Always consider the risks of continued antibiotic avoidance, particularly when a narrow-spectrum antibiotic is the antibiotic of choice for a documented bacterial infection, in relation to the risks of reexposure when managing antibiotic intolerance.

REFERENCES

1. Fleming-Dutra KE, Hersh AL, Shapiro DJ, Bartoces M, Enns EA, File TM Jr, et al. Prevalence of inappropriate antibiotic prescriptions among US ambulatory care visits, 2010-2011. *JAMA* 2016;315:1864-73.

2. European Centre for Disease Prevention and Control. Data and reports: Antimicrobial resistance and consumption. Available from: <http://ecdc.europa.eu/en/eaad/antibiotics-get-informed/antibiotics-resistance-consumption/pages/data-reports.aspx>. Accessed November 18, 2016.
3. Kaye KS, Poque JM. Infections caused by resistant gram-negative bacteria: epidemiology and management. *Pharmacotherapy* 2015;35:949-62.
4. Barlam TF, Cosgrove SE, Abbo LM, MacDougall C, Schuetz AN, Septimus EJ, et al. Implementing an antibiotic stewardship program: guidelines by the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America. *Clin Infect Dis* 2016;62:e51-77.
5. Jeffres MN, Narayanan PP, Shuster JE, Schramm GE. Consequences of avoiding β -lactams in patients with β -lactam allergies. *J Allergy Clin Immunol* 2016;137:1148-53.
6. Joint Task Force on Practice Parameters; American Academy of Allergy, Asthma, and Immunology; American College of Allergy, Asthma, and Immunology; Joint Council of Allergy, Asthma, and Immunology. Drug allergy: an updated practice parameter. *Ann Allergy Asthma Immunol* 2010;105:259-73.
7. Brockow K, Garvey LH, Aberer W, Atanaskovic-Markovic M, Barbaud A, Bilo MB, et al. Skin test concentrations for systemically administered drugs – an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy* 2013;68:702-12.
8. Mayorga C, Celik G, Rouzair P, Whitaker P, Bonadonna P, Cernadas JR, et al. In vitro tests for drug hypersensitivity reactions: an ENDA/EAACI Drug Allergy Interest Group position paper. *Allergy* 2016;71:1103-34.
9. Blumenthal KG, Park MA, Macy EM. Redesigning the allergy module of the electronic health record. *Ann Allergy Asthma Immunol* 2016;117:126-31.
10. Gerace KS, Karlin E, Mckinnon E, Phillips E. Varying penicillin allergy testing practices in the United States: a time for consensus. *J Allergy Clin Immunol Pract* 2015;3:791-3.
11. Bamanikar A. A review of drug allergies: diagnosis and management. *EMJ Allergy Immunol* 2016;1:52-7.
12. Macy E. Penicillin allergy: optimizing diagnostic protocols, public health implications, and future research needs. *Curr Allergy Asthma Rep* 2016;16:4.
13. Topaz M, Goss F, Blumenthal K, Lai K, Seger DL, Slight SP, et al. Toward improved drug allergy alerts: multidisciplinary expert recommendations. *J Am Med Inform Assoc* 2016;23:601-8.
14. Macy E, Poon KWT. Self-reported antibiotic allergy incidence and prevalence: age and sex effects. *Am J Med* 2009;122. 778.e1-778.e7.
15. Macy E, Ho NJ. Multiple drug intolerance syndrome: prevalence, clinical characteristics, and management. *Ann Allergy Asthma Immunol* 2012;108:88-93.
16. Zhou L, Dhopeswarkar N, Blumenthal KG, Goss F, Topaz M, Slight SP, et al. Drug allergies documented in electronic health records or a large healthcare system. *Allergy* 2016;71:1305-13.
17. Salden OAE, Rockmann H, Verheij TJM, Broekhuizen BDL. Diagnosis of allergy against beta-lactams in primary care: prevalence and diagnostic criteria. *Family Pract* 2015;32:257-62.
18. Anderson GD. Gender differences in pharmacological response. *Int Rev Neurobiol* 2008;83:1-10.
19. Macy E, Mangat R, Burchette RJ. Penicillin skin testing in advance of need: multiyear follow-up in 568 test result-negative subjects exposed to oral penicillins. *J Allergy Clin Immunol* 2003;111:1111-5.
20. Terpening C. Should antibiotic allergy alerts be alarming? *Southern Med J* 2016;109:648-52.
21. Romano A, Torres MJ, Castells M, Sanz ML, Blanca M. Diagnosis and management of drug hypersensitivity reactions. *J Allergy Clin Immunol* 2011;127:S67-73.
22. Demoly P, Adkinson NF, Brockow K, Castells M, Chiriac AM, Greenberger PA, et al. International consensus on drug allergy. *Allergy* 2014;69:420-37.
23. Bircher AJ, Scherer Hofmeier K. Drug hypersensitivity reactions: inconsistency in the use of the classification of immediate and nonimmediate reactions. *J Allergy Clin Immunol* 2012;129:263-4. author reply.
24. Pichler WJ. Delayed drug hypersensitivity reactions. *Ann Intern Med* 2003;139:683-93.
25. Renaudin JM, Beaudouin E, Ponvert C, Demoly P, Moneret-Vautrin DA. Severe drug-induced anaphylaxis: analysis of 333 cases recorded by the Allergy Vigilance Network from 2002 to 2010. *Allergy* 2013;68:929-37.
26. Macy E, Schatz M, Lin CK, Poon KW. The falling rate of positive penicillin skin tests from 1995 to 2007. *Perm J* 2009;13:12-9.
27. Rosenfield L, Kalicinsky C, Warrington R. A retrospective comparison of false negative skin test rates in penicillin allergy, using penicilloyl-poly-lysine and minor determinants or penicillin G, followed by open challenge. *Allergy Asthma Clin Immunol* 2015;11:34.
28. Romano A, Blanca M, Torres MJ, Bircher A, Aberer W, Brockow K, et al. Diagnosis of nonimmediate reactions to beta-lactam antibiotics. *Allergy* 2004;59:1153-60.
29. Cheng CY, Su SC, Cheen CH, Cheen WL, Deng ST, Chung WH. HLA associations and clinical implications in T-cell mediated drug hypersensitivity reactions: an updated review. *J Immunol Res* 2014;2014:565320.
30. Ricklin D, Reis ES, Lambris JD. Complement in disease: a defense system turning offensive. *Nat Rev Nephrol* 2016;12:383-401.
31. Kracker S, Durandy A. Insights into B cell specific process of immunoglobulin class switch recombination. *Immunol Lett* 2011;138:97-103.
32. Blumenthal KG, Saff RR, Banerji A. Evaluation and management of a patient with multiple drug allergies. *Allergy Asthma Proc* 2014;35:197-203.
33. Macy E. Penicillin allergy: optimizing diagnostic protocols, public health implications, and future research needs. *Curr Opin Allergy Clin Immunol* 2015;15:308-13.
34. Macy E, Contreras R. Health care use and serious infection prevalence associated with penicillin "allergy" in hospitalized patients: a cohort study. *J Allergy Clin Immunol* 2014;133:790-6.
35. Caubet JC, Frossard C, Fellay B, Eigenmann PA. Skin tests and in vitro allergy tests have a poor diagnostic value for benign skin rashes due to β -lactams in children. *Pediatr Allergy Immunol* 2015;26:80-2.
36. Romano A, Gaeta F, Poves MF, Valluzzi RL. Cross-reactivity among beta-lactams. *Curr Allergy Asthma Rep* 2016;16:24.
37. Romano A, Bousquet-Rouanet L, Viola M, Gaeta F, Demoly P, Bousquet PJ. Benzylpenicillin skin testing is still important in diagnosing immediate hypersensitivity reactions to penicillins. *Allergy* 2009;64:249-53.
38. Torres MJ, Romano A, Mayorga C, Moya MC, Guzman AE, Reche M, et al. Diagnostic evaluation of a large group of patients with immediate allergy to penicillins: the role of skin testing. *Allergy* 2001;56:850-6.
39. Confino-Cohen R, Rosman Y, Lachover I, Meir Shafir K, Goldberg A. The importance of amoxicillin and amoxicillin-clavulanate determinants in the diagnosis of immediate allergic reactions to β -lactams. *Int Arch Allergy Immunol* 2016;170:62-6.
40. Bousquet PJ, Pipet A, Bousquet-Rouanet L, Demoly P. Oral challenges are needed in the diagnosis of beta-lactam hypersensitivity. *Clin Exp Allergy* 2008;38:185-90.
41. Mill C, Primeau MN, Medoff E, Lejtenyi C, O'Keefe A, Netchiporouk E, et al. Assessing the diagnostic properties of a graded oral provocative challenge for the diagnosis of immediate and nonimmediate reactions to amoxicillin in children. *JAMA Pediatr* 2016;170:e160033.
42. Macy E, Ngor EW. Safely diagnosing clinically significant penicillin allergy using only penicilloyl-poly-lysine, penicillin, and oral amoxicillin. *J Allergy Clin Immunol Pract* 2013;1:258-63.
43. Schnyder B, Pichler WJ. Skin and laboratory tests in amoxicillin- and penicillin-induced morbilliform skin eruption. *Clin Exp Allergy* 2000;30:590-5.
44. Barbaud AM, Bene MC, Schmutz JL, Ehlinger A, Weber M, Faure GC. Role of delayed cellular hypersensitivity and adhesion molecules in amoxicillin-induced morbilliform rashes. *Arch Dermatol* 1997;133:481-6.
45. Harandian F, Pham D, Ben-Shoshan M. Positive penicillin allergy testing results: a systematic review and meta-analysis of papers published from 2010 through 2015. *Postgrad Med* 2016;128:557-62.
46. Solensky R, Khan DA. Evaluation of antibiotic allergy: the role of skin tests and drug challenges. *Curr Allergy Asthma Rep* 2014;14:459.
47. Macy E, Goldberg B, Poon KY. Use of commercial anti-penicillin IgE fluorometric enzyme immunoassays to diagnose penicillin allergy. *Ann Allergy Asthma Immunol* 2010;105:136-41.
48. Johansson SG, Adedoyin J, van Hage M, Gronneberg R, Nopp A. False-positive penicillin immunoassay: an unnoticed common problem. *J Allergy Clin Immunol* 2013;132:235-7.
49. Pastorello EA, Stafylaraki C, Mirone C, Preziosi D, Aversano MG, Macheri A, et al. Anti-amoxicillin immunoglobulin E, histamine-2 receptor antagonist therapy and mast cell activation syndrome are risk factors for amoxicillin anaphylaxis. *Int Arch Allergy Immunol* 2015;166:280-6.
50. Cabanas R, Calderon O, Ramirez E, Fiander A, Prior N, Caballero T, et al. Piperacillin-induced DRESS: distinguishing features in a clinical and allergy study of 8 patients. *J Investig Allergol Clin Immunol* 2014;24:425-30.
51. Zawodniak A, Lochmatter P, Beeler A, Pichler WJ. Cross-reactivity in drug hypersensitivity reactions to sulfasalazine and sulfamethoxazole. *Int Arch Allergy Immunol* 2010;153:152-6.
52. Schnyder B, Pichler WJ. Allergy to sulfonamides. *J Allergy Clin Immunol* 2013;131:256-257.e1-5.

53. Gruchalla RS. 10. Drug allergy. *J Allergy Clin Immunol* 2003;111:S548-59.
54. Watkins S, Pichler WJ. Sulfamethoxazole induces a switch mechanism in T cell receptors containing TCRV β 20-1, altering pHLA recognition. *PLoS One* 2013;8:e76211.
55. Chantachaeng W, Chularojanamontri L, Kulthanan K, Jongjareanprasert K, Dhana N. Cutaneous adverse reactions to sulfonamide antibiotics. *Asian Pac J Allergy Immunol* 2011;29:284-9.
56. Lin D, Li WK, Rieder MJ. Cotrimoxazole for prophylaxis or treatment of opportunistic infections of HIV/AIDS in patients with previous history of hypersensitivity to cotrimoxazole. *Cochrane Database Syst Rev* 2007;18:CD005646.
57. Kuyucu S, Mori F, Atanaskovic-Markovic M, Caubet JC, Terreehorst I, Gomes E, et al. Pediatric Task Force of EAACI Drug Allergy Interest Group. Hypersensitivity reactions to non-beta-lactam antibiotics in children: an extensive review. *Pediatr Allergy Immunol* 2014;25:534-43.
58. Pyle RC, Butterfield JH, Volcheck GW, Podjasek JC, Rank MA, Li JT, et al. Successful outpatient graded administration of trimethoprim-sulfamethoxazole in patients without HIV and with a history of sulfonamide adverse drug reaction. *J Allergy Clin Immunol Pract* 2014;2:52-8.
59. Beumont MG, Graziani A, Ubel PA, MacGregor RR. Safety of dapsone as *Pneumocystis carinii* pneumonia prophylaxis in human immunodeficiency virus-infected patients with allergy to trimethoprim/sulfamethoxazole. *Am J Med* 1996;100:611-6.
60. Bonfanti P, Pusterla L, Parazzini F, Libanore M, Cagni AE, Franzetti M, et al. The effectiveness of desensitization versus rechallenge treatment in HIV-positive patients with previous hypersensitivity to TMP-SMX: a randomized multicentric study. C.I.S.A.I. Group. *Biomed Pharmacother* 2000;54:45-9.
61. Leoung GS, Stanford JF, Giordano MF, Stein A, Torres RA, Giffen CA, et al. Trimethoprim-sulfamethoxazole (TMP-SMZ) dose escalation versus direct rechallenge for *Pneumocystis carinii* pneumonia prophylaxis in human immunodeficiency virus-infected patients with previous adverse reaction to TMP-SMZ. *J Infect Dis* 2001;184:992-7.
62. Araujo L, Demoly P. Macrolides allergy. *Curr Pharmaceut Design* 2008;14:2840-62.
63. Hicks LA, Taylor TH Jr, Hunkler RJ. U.S. outpatient antibiotic prescribing, 2010. *N Engl J Med* 2013;368:1461-2.
64. Seitz CS, Brocker EB, Trautmann A. Suspicion of macrolide allergy after treatment on infectious diseases including *Helicobacter pylori*: results of allergological testing. *Allergol Immunopathol (Madrid)* 2011;39:193-9.
65. Cavkaytar O, Karaatmaca B, Yilmaz EA, Sekerel BE, Soyer O. Testing for clarithromycin hypersensitivity: a diagnostic challenge in childhood. *J Allergy Clin Immunol Pract* 2016;4:330-2.
66. Mori F, Pecorari L, Pantano S, Rossi ME, Pucci N, De Martino M, et al. Azithromycin anaphylaxis in children. *Int J Immunopathol Pharmacol* 2014;27:121-6.
67. Benahmed S, Scaramuzza C, Messaad D, Sahla H, Demoly P. The accuracy of the diagnosis of suspected macrolide antibiotic hypersensitivity: results of a single-blinded trial. *Allergy* 2004;59:1130-3.
68. Macy E, Contreras R. Adverse reactions associated with oral and parenteral use of cephalosporins: a retrospective population-based analysis. *J Allergy Clin Immunol* 2015;135:745-752.e5.
69. Romano A, Gaeta F, Valluzzi RL, Caruso C, Rumi G, Bousquet PJ. IgE-mediated hypersensitivity to cephalosporins: cross-reactivity and tolerability of penicillins, monobactams, and carbapenems. *J Allergy Clin Immunol* 2010;126:994-9.
70. Macy E. Penicillin and beta-lactam allergy: epidemiology and diagnosis. *Curr Allergy Asthma Rep* 2014;14:476.
71. Uyttebroek AP, Decuyper IL, Bridts CH, Romano A, Hagendorens MM, Ebo DG, et al. Cefazolin hypersensitivity: toward optimized diagnosis. *J Allergy Clin Immunol Pract* 2016;4:1232-6.
72. Dickson SD, Salazar KC. Diagnosis and management of immediate hypersensitivity reactions to cephalosporins. *Clin Rev Allergy Immunol* 2013;45:131-42.
73. Drucker AM, Rosen CF. Drug-induced photosensitivity: culprit drugs, management and prevention. *Drug Saf* 2011;34:821-37.
74. Lobera T, Audicana MT, Alarcón E, Longo N, Navarro B, Muñoz D. Allergy to quinolones: low cross-reactivity to levofloxacin. *J Investig Allergol Clin Immunol* 2010;20:607-11.
75. McNeil BD, Pundir P, Meeker S, Han L, Udem BJ, Kulka M, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature* 2015;519:237-41.
76. Uyttebroek AP, Sabato V, Bridts CH, De Clerck LS, Ebo DG. Moxifloxacin hypersensitivity: uselessness of skin testing. *J Allergy Clin Immunol Pract* 2015;3:443-5.
77. Mori K, Maru C, Takasuna K, Furuhashi K. Mechanism of histamine release induced by levofloxacin, a fluoroquinolone antibacterial agent. *Eur J Pharmacol* 2000;394:51-5.
78. Aspinall SL, Good CB, Jiang R, McCarren M, Dong D, Cunningham FE. Severe dysglycemia with the fluoroquinolones: a class effect? *Clin Infect Dis* 2009;59:402-8.
79. Kalegasioglu F, Olcay E. Fluoroquinolone-induced tendinopathy: etiology and preventive measures. *Tohoku J Exp Med* 2012;226:251-8.
80. Paiva LA, Wright PJ, Koff RS. Long-term hepatic memory for hypersensitivity to nitrofurantoin. *Am J Gastroenterol* 1992;87:891-3.
81. Thornhill MH, Dayer MJ, Prendergast B, Baddour LM, Jones S, Lockart PB. Incidence and nature of adverse reactions to antibiotics used as endocarditis prophylaxis. *J Antimicrob Chemother* 2015;70:2382-8.
82. Pereira N, Canelas MM, Santiago F, Brites MM, Goncalo M. Value of patch tests in clindamycin-related drug eruptions. *Contact Dermatitis* 2011;65:202-7.
83. Pearlman MD, Yashar C, Ernst S, Solomon W. An incremental dosing protocol for women with severe vaginal trichomoniasis and adverse reactions to metronidazole. *Am J Obstet Gynecol* 1996;174:934-6.
84. Moss RB. Sensitization to aztreonam and cross-reactivity with other beta-lactam antibiotics in high-risk patients with cystic fibrosis. *J Allergy Clin Immunol* 1991;87:78-88.
85. Vega JM, Blanca M, Garcia JJ, Miranda A, Carmona MJ, Garcia A, et al. Tolerance to aztreonam in patients allergic to beta-lactam antibiotics. *Allergy* 1991;46:196-202.
86. Atanaskovic-Markovic M, Gaeta F, Medjo B, Viola M, Nestorovic B, Romano A. Tolerability of meropenem in children with IgE-mediated hypersensitivity to penicillins. *Allergy* 2008;63:237-40.
87. Atanaskovic-Markovic M, Gaeta F, Gavrovic-Jankulovic M, Velickovic TC, Valluzzi RL, Romano A. Tolerability of imipenem in children with IgE-mediated hypersensitivity to penicillins. *J Allergy Clin Immunol* 2009;124:167-9.
88. Romano A, Gaeta F, Valluzzi RL, Alonzi C, Maggioletti M, Zaffiro A, et al. Absence of cross-reactivity to carbapenems in patients with delayed hypersensitivity to penicillins. *Allergy* 2013;68:1618-21.
89. Sanchez-Morillas L, Perez-Ezquerro PR, Reano-Martos M, Laguna-Martinez JJ, Sanz ML, Martinez LM. Selective allergic reactions to clavulanic acid: a report of 9 cases. *J Allergy Clin Immunol* 2010;126:177-9.
90. Sánchez-Borges M, Thong B, Blanca M, Ensina LF, González-Díaz S, Greenberger PA, et al. Hypersensitivity reactions to non-beta-lactam antimicrobial agents, a statement of the WAO special committee on drug allergy. *World Allergy Organ J* 2013;6:18.
91. Liippo J, Lammintausta K. Positive patch test reactions to gentamicin show sensitization to aminoglycosides from topical therapies, bone cements, and from systemic medication. *Contact Dermatitis* 2008;59:268-72.
92. Minhas JS, Wickner PG, Long AA, Banerji A, Blumenthal KG. Immune-mediated reactions to vancomycin: a systematic case review and analysis. *Ann Allergy Asthma Immunol* 2016;116:544-53.
93. Bernedo N, Gonzalez I, Gastaminza G, Audicana M, Fernandez E, Munoz D. Positive patch test in vancomycin allergy. *Contact Dermatitis* 2001;45:43.
94. Otani IM, Kuhlen JL Jr, Blumenthal KG, Guyer A, Banerji A. A role for vancomycin epicutaneous skin testing in the evaluation of perioperative anaphylaxis. *J Allergy Clin Immunol Pract* 2015;3:984-5.
95. Kao L, Rajan J, Roy L, Kavosh E, Khan DA. Adverse reactions during drug challenges: a single US institution's experience. *Ann Allergy Asthma Immunol* 2013;110:86-91.e1.
96. Iammatteo M, Blumenthal KG, Saff R, Long AA, Banerji A. Safety and outcomes of test doses for the evaluation of adverse drug reactions: a 5-year retrospective review. *J Allergy Clin Immunol Pract* 2014;2:768-74.
97. Demoly P, Romano A, Botelho C, Bousquet-Rouanet L, Gaeta F, Silva R, et al. Determining the negative predictive value of provocation tests with beta-lactams. *Allergy* 2010;65:327-32.
98. Aberer W, Bircher A, Romano A, Blanca M, Campi P, Fernandez J, et al. Drug provocation testing in the diagnosis of drug hypersensitivity reactions: general considerations. *Allergy* 2003;58:854-63.
99. Romano A, Gaeta F, Valluzzi RL, Maggioletti M, Zaffiro A, Caruso C, et al. IgE-mediated hypersensitivity to cephalosporins: cross-reactivity and tolerability of alternative cephalosporins. *J Allergy Clin Immunol* 2015;136:685-691.e3.
100. Ponvert C, Perrin Y, Bados-Albiero A, Le Bourgeois M, Karila C, Delacourt C, et al. Allergy to beta-lactam antibiotics in children: results of a

- 20-year study based on clinical history, skin and challenge tests. *Pediatr Allergy Immunol* 2011;22:411-8.
101. Kearns GL, Wheeler JG, Rieder MJ, Reid J. Serum sickness-like reaction to cefaclor: lack of in vitro cross-reactivity with loracarbef. *Clin Pharmacol Ther* 1998;93:686-93.
 102. Khan DA. Treating patients with multiple drug allergies. *Ann Allergy Asthma Immunol* 2013;110:2-6.
 103. Peck SM, Siegal S, Glick AW, Kurtin A, Bergamini R. Clinical problems in penicillin sensitivity. *JAMA* 1948;138:631-40.
 104. Sullivan TJ, Yecies LD, Shatz GS, Parker CW, Wedner HJ. Desensitization of patients allergic to penicillin using orally administered beta-lactam antibiotics. *J Allergy Clin Immunol* 1982;69:275-82.
 105. Wendel GD Jr, Stark BJ, Jamison RB, Molina RD, Sullivan TJ. Penicillin allergy and desensitization in serious infections during pregnancy. *N Engl J Med* 1985;312:1229-32.
 106. Castells M. Rapid desensitization for hypersensitivity reactions to medications. *Immunol Allergy Clin North Am* 2009;29:585-606.
 107. Scherer K, Brockow K, Aberer W, Gooi JH, Demoly P, Romano A, et al. Desensitization in delayed drug hypersensitivity reactions – an EAACI position paper of the Drug Allergy Interest Group. *Allergy* 2013;68:844-52.
 108. Cernadas JR, Brockow K, Romano A, Aberer W, Torres MJ, Bircher A, et al. General considerations on rapid desensitization for drug hypersensitivity—a consensus statement. *Allergy* 2010;65:1357-66.